

### Manual

# MagnifiQ™ 1 Pathogen instant kit

Kit for automated, magnetic isolation of DNA and RNA of pathogens in the strip format. Contains ready-to-use, reagent-filled stripes and all necessary consumables.

REF	size	compatible devices *
607A-1V-32	32 isolations	Auto-Pure Mini Auto-Pure S32
607A-1V-160	160 isolations	Auto-Pure Mini Auto-Pure S32

#### \* Compatible devices

The kit has been tested with specific Allsheng brand isolation devices. This does not preclude it from working with other devices. If your device is not listed, please contact us at info@aabiot.com.

For in vitro diagnostics use.





A&A Biotechnology, Strzelca 40, 80-299 Gdańsk, Poland

A&A Biotechnology provides a guarantee on this product.

The company does not guarantee the correct performance of this kit in the event of:

- not adhering to the supplied protocol
- use of not recommended equipment or materials
- use of other reagents than recommended or which are not a component of the product
- use of expired or improperly stored product or its components.

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### **Advantages**

- Requires only a few minutes of manual work while adding the samples. The rest of the procedure is carried
  out in an automatic extraction device.
- It does not require the initial preparation of buffers. Just add prepared samples to the plate and after about 30 min you get extracted material.
- Universal extraction of both DNA and RNA.

### Sample types

sample size
up to 20 mg
up to 200 μl
1 pcs

### **Specification**

protocol time	~ 30 min
elution volume	100 μΙ
elution solution	Tris buffer (10 mM, pH 8.5)
binding capacity	30 μg DNA/RNA
downstream applications	qPCR, RT-qPCR, sequencing

### Description

MagnifiQ<sup>™</sup> 1 Pathogen instant kit is designed for RNA and DNA isolation from viruses and Gram(-) bacteria. The isolated material is perfect for further analyzes and tests by qPCR and RT-PCR methods and for sequencing.

The Magnifi $Q^{\mathbf{m}}$  product series is based on the automated isolation of nucleic acids with use of magnetic beads. This method significantly shortens working time and reduces risk of mistakes in comparison to manual methods.

#### **Contents**

	607A-1V-32		607A-1V-160		
component	quantity	cat#	quantity	cat#	storage
XS-G - extraction strip	4 x 8 pcs	K-S1V22XG	20 x 8 pcs	K-S1V22XG	15-25 ℃
LTE 2X buffer	8 ml	K-LTE2X-8	35 ml	K-LTE2X-35	15-25 ℃
Proteinase K	1,1 ml	K-PRK-11A	4 x 1,1 ml	K-PRK-11A	2-8 °C*
tip comb 8	16 pcs	K-C8U-16	2 x 40 pcs	K-C8U-40	15-25 ℃

<sup>\*</sup> Proteinase K can be stored at 15-25 °C for up to 12 months.

### Additional equipment and reagents

#### **Necessary**

- pipette
- pipette tips
- 1.5 ml Eppendorf tubes with lock (sample lysis)

#### **Optional**

- vortex
- sterile water. Tris buffer. PBS buffer

### **Material preparation**

#### **Animal tissue**

- Homogenize up to 20 mg of animal tissue in the PBS or Tris buffer.
   Centrifuge the sample for 1 min at 500 RPM.
   Transfer 200 μl of the supernatant to 1.5 ml Eppendorf tube with lock.
   Add 200 μl of LTE 2X buffer and 20 μl of Proteinase K.
  - 5. Vortex the sample for **10 s**.
  - 6. Follow point 1. of the Extraction Protocol.

### Blood (fresh or frozen, plasma, serum), body fluids

- 1. Transfer 200 μl of the sample to 1.5 ml Eppendorf tube with lock.
- 2. Add 200 μl of LTE 2X and 20 μl of Proteinase K.
- Vortex the sample for 10 s and incubate for 10 min at 50 °C with shaking.
   Note. If automatic shaking is not available, mix the samples by inverting the tubes a few times.
- 4. Centrifuge for 20 s at 10 000 RPM.

5.

Attention. In the Extraction Protocol, use the supernatant as the sample.

Follow point 1. of the Extraction Protocol.

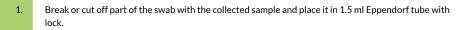
### Swabs with transport medium

No additional material preparation is required.



Follow point 1. of the Extraction Protocol.

### Dry swabs



Note. The portion of the swab with the collected sample should fit completely into the tube.

2. Add 500 μl of sterile water, Tris buffer or PBS buffer.

Note. Part of the swab with the sample should be completely immersed in the buffer.

- 3. Leave at room temperature for 10 min.
- 4. Vortex for 10 s.
- 5. Transfer 200 μl of supernatant to new 1.5 ml Eppendorf tube with lock.
- 6. Add 200 μl of LTE 2X buffer and 20 μl of Proteinase K.
- 7. Follow point 1. of the Extraction Protocol.

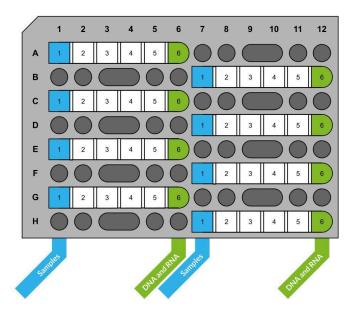
## **Protocol**

### **Protocol files**

device	protocol name	protocol file	ins	tallation
Auto-Pure Mini M		aabiot.com/protocols/magnifig /MI/MO-UNI-MI.txt	1.	Create folder "items" on a USB drive and copy the protocol file to it.
			2.	Insert the USB drive into a USB slot in the device.
	MQ-UNI-MI		3.	On a device screen, go to Settings > System > Transfer > Import.
			4.	Select the protocol and tap "Import".
Auto-Pure Mini (QR code)	MQ-UNI-MI		1. 2.	On a device screen, go to Run > \( \begin{align*} \operatorname \operat
Auto-Pure S32	MQ_UNI_\$32	aabiot.com/protocols/magnifiq /S32/MO_UNI_S32.txt	1.	Create folder "im_export_protocols" on a USB drive and copy the protocol file to it.
			2.	Insert the USB drive into a USB slot in the device.
			3.	On a device screen, go to Protocols >Import.
			4.	Select the protocol and tap "Import".

#### **Extraction Protocol**

1. Place **XS-G** stripes in the rack(s).



2. Remove the foil from the **XS-G** stipes.

Note: It is important to do this carefully so as not to mix buffers from different wells.

Note: If possible, briefly spin before removing the foil.

3. Add 400 µl of sample to the well 1 (first from the left) on the XS-G strip.

Note. The wells are numbered on the side of the strip.

- 4. Place the racks(s) in the extraction device.
- 5. Place the appropriate number of **tip combs 8** in the extraction device.
- 6. Run the protocol on your device.
- 7. After the run is over, first remove the combs and then the racks from the extraction device and transfer the purified DNA / RNA located in well 6 (first from the right) on the XS-G strip to a sterile tube.

Note. For longer storage of extracted material, transfer it from the plate to appropriate tubes and store at  $4 \,^{\circ}\text{C}$  for DNA or  $-70 \,^{\circ}\text{C}$  for RNA.

# **Troubleshooting**

Problem	Possible cause	Suggested solution		
No nucleic acid at all.	Material was not processed.	Please ensure the sample lysate was added to the extraction strip. Inspect the volumes of the respective sample strip wells. In case of uncertainty repeat the procedure with new sample material.		
	Automated extraction protocol was not started.	Please check the position of the beads in the strip. If the beads were not transferred from their original position START the protocol for nucleic acid extraction.		
	Initial sample material was partially degraded.	Please repeat the isolation with the same and different sample type to double check the sample quality.		
Low RNA yield.	Elution well was contaminated with exogenous RNAse.	Please repeat the isolation with the fresh portion of elution reagent to exclude the RNAse contamination.		
Low DNA yield	Extraction of DNA during	To obtain higher yields of DNA, the incubation time in lysis buffer can be prolonged (usually up to 20 min).		
	sample lysis was not sufficient.	For sample that the lysis is supported by Proteinase K digestion please ensure the Proteinase K was added or increase the volume of enzyme up to $40  \mu l$ per sample.		
	Sample contains too much RNA.	Add 10–20 $\mu$ I RNase A solution to the lysis buffer before heat incubation. If this is not successful, add the enzyme to the cleared lysate and incubate for 30 min at 37 °C.		
	Suboptimal elution.	The DNA can be either eluted in higher or lower volumes (40-100 µL) or by prolonging the elution step up. Check your automated extractor programme and if needed decrease the elution volume in the elution plate/row.		
		Note that it is advantageous to perform the elution pre-heated to 65-70 $^{\circ}$ C. Check your automated extractor programme.		
Degraded DNA	Sample was contaminated with DNAse.	Check working area and pipettes for DNAse content. Use cleaning product to remove any enzymatic activity contamination.		
	Sample dependent problem.	Highly processed samples may be responsible for impossibility to extract high molecular weight DNA.		
Low DNA / RNA quality	Sample contains DNA-degrading contaminants (e.g., phenolic compounds, metabolites)	Investigate if repeating the wash 1 or EtOH 80% wash would improve the quality of eluate.		

### Safety information

### Proteinase K



H315 Causes skin irritation.

H319 Causes serious eye irritation.

H334 May cause allergy or asthma symptoms or breathing difficulties if inhaled.

H335 May cause respiratory irritation.

P261 Avoid breathing dust.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,

if present and easy to do. Continue rinsing.

 $P342 + P311\ If\ experiencing\ respiratory\ symptoms\ call\ a\ Poison\ Center\ or\ doctor/physician.$ 



WARNING

#### LTE 2X

H302 Harmful if swallowed.

H315 Causes skin irritation.

H319 Causes serious eye irritation.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,

if present and easy to do. Continue rinsing.

#### XS-G extraction strip











H225 Highly flammable liquid and vapor.

H302+H312+H332 Harmful if swallowed, in contact with skin or if inhaled.

H314 Causes severe skin burns and eye damage.

H412 Harmful to aquatic life with long lasting effects.

P210 Keep away from heat, hot surfaces, sparks, open flames and other ignition sources. No smoking. P273 Avoid release to the environment.

P280 Wear protective gloves/protective clothing/eye protection/face protection/hearing protection.

P301+P312+P330 If swallowed: Call a poison center/doctor/ if you feel unwell.

P303+P361+P353 If on skin (or hair): Take off immediately all contaminated clothing. Rinse skin with water or shower.

P304+P340 If inhaled: Remove person to fresh air and keep comfortable for breathing.

P305+P351+P338 If in eyes: rinse cautiously with water for several minutes. Remove contact lenses,

if present and easy to do. Continue rinsing.

## **Explanation of symbols used**

symbol	symbol meaning	symbol	symbol meaning
IVD	Indicates the <i>In vitro</i> diagnostics medical device	REF	Indicates the manufacturer's catalogue number
***	Indicates the medical device manufacturer	[]i	Indicates the need for the user to consult the instructions for use
LOT	Indicates the manufacturer's batch code or lot can be identified	Ţ	Indicates the need for the user to consult the instructions for use for important information such as warnings and cautions
	Indicates the date after which the medical device is not to be used	1	Indicates the temperature limits to which the medial device can be safely exposed



A&A Biotechnology, Strzelca 40, 80-299 Gdańsk tel. 883 323 761, 600 776 268 info@aabiot.com, www.aabiot.com

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